



## On-field evaluation of an ultra-rapid fluorescence immunoassay as a frontline test for SARS-CoV-2 diagnostic

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### ABSTRACT

**Background:** Viral RNA amplification by real-time RT-PCR still represents the gold standard for the detection of SARS-CoV-2, but the development of rapid, reliable and easy-to-perform diagnostic methods is crucial for public health, because of the need of shortening the time of result-reporting with a cost-efficient approach.

**Objectives:** The aim of our research was to assess the performance of FREN<sup>TM</sup> COVID-19 Ag assay (NanoEntek, South Korea) as a ultra-rapid frontline test for SARS-CoV-2 identification, in comparison with RT-PCR and another COVID-19 antigen fluorescence immunoassay (FIA).

**Study design:** The qualitative FIA FREN<sup>TM</sup> test, designed to detect within 3 min the Nucleocapsid protein of SARS-CoV-2, was evaluated using nasopharyngeal swabs in Universal Transport Medium (UTM<sup>TM</sup>, Copan Diagnostics Inc, US) from suspected COVID-19 cases who accessed the Emergency Room of the Ospedale Policlinico San Martino, Genoa, Liguria, Northwest Italy. Diagnostic accuracy was determined in comparison with SARS-CoV-2 RT-PCR and STANDARD F<sup>TM</sup> COVID-19 Ag FIA test (SD BIOSENSOR Inc., Republic of Korea).

**Results:** In November 2020, 110 nasopharyngeal samples were collected consecutively; 60 resulted RT-PCR positive. With respect to RT-PCR results, sensitivity and specificity of FREN<sup>TM</sup> COVID-19 Ag test were 93.3 % (95 % CI: 83.8–98.2) and 100 % (95 % CI: 92.9–100), respectively.

FREN<sup>TM</sup> and STANDARD F<sup>TM</sup> COVID-19 Ag FIA assays showed a concordance of 96.4 % (Cohen's  $k = 0.93$ , 95 % CI: 0.86–0.99).

**Conclusions:** FREN<sup>TM</sup> FIA test showed high sensitivity and specificity in nasopharyngeal swabs. The assay has the potential to become an important tool for an ultra-rapid identification of SARS-CoV-2 infection, particularly in situations with limited access to molecular diagnostics.

## 1. Background

According to the World Health Organization (WHO), Real Time reverse transcription polymerase chain reaction (RT-qPCR) on respiratory samples (nasopharyngeal swabs) is the current recommended laboratory method to diagnose SARS-CoV-2 acute infection, the cause of 2019 coronavirus disease (COVID-19) (Corman et al., 2020; OMS, 2020).

In 2020, the increasing molecular analysis demand due to the COVID-19 pandemic raised several critical issues such as requirement of special equipment, laboratory reagents and skilled staff.

Therefore, looking for alternative diagnostic solutions to implement

a molecular screening strategy extended to a large number of subjects and to counter the SARS-CoV-2 spread has become a priority for the health care systems (Mak et al., 2020).

In the last months, several easy to perform rapid antigen detection tests were developed and recommended in some countries as first line laboratory strategy for COVID-19 diagnostic (Schoy et al., 2020; Porte et al., 2020).

## 2. Objectives

The aim of the present research was to assess the performance of FREN<sup>TM</sup> COVID-19 Ag assay (NanoEntek, South Korea) as a frontline

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test for SARS-CoV-2 identification in comparison to the available molecular techniques and to the STANDARD F COVID-19 Ag FIA test (SD BIOSENSOR Inc., Republic of Korea), a rapid fluorescent immunoassay test currently used in clinical practice (Cerutti et al., 2020).

### 3. Study design

In November 2020, at the regional reference laboratory for COVID-19 diagnostic located into Ospedale Policlinico San Martino, Hygiene Unit, Genoa, Liguria, Northwest Italy, 110 nasopharyngeal samples from 110 patients who accessed the Emergency Room of the hospital with symptoms attributable to SARS-CoV-2 infection were consecutively collected. Swabs were carried out using a flocked probe and eluted in Universal Transport Medium (UTM™, Copan Diagnostics Inc, US). RT-qPCR, FRENDS™ COVID-19 Ag and STANDARD F COVID-19 Ag FIA tests were conducted within 8 h from the swabs' arrival at the laboratory.

Each respiratory swab was set up for PCR using the extraction-free method on Nimbus IVD, (Seegene, South Korea) using the Allplex™ SARS-CoV-2 Assay kit (Seegene, South Korea), according to the manufacturer's instructions. The obtained material was tested for the identification of SARS-CoV-2 through a one-step multiplex RT-qPCR on Biorad CFX96™ thermal cycler. It targeted the nucleoprotein region (N), the RNA-dependent RNA-polymerase region (RdRp), the Spike protein (S) and the Envelop region (E), respectively. Samples showing a cycle threshold (Ct) value <35 for all the target genes were considered positive.

Positive swabs were tested in parallel with FRENDS™ COVID-19 Ag rapid diagnostic test for SARS-CoV-2 infection. It was a qualitative fluorescence immunoassay (FIA) designed to detect in nasopharyngeal swabs within 3 min the Nucleocapsid protein (N) of SARS-CoV-2 and intended for use with FRENDS™ system (NanoEntek, South Korea). Results were displayed as positive or negative for COVID-19 antigen and a numeric value was provided ( $\geq 1.00$  for positive samples).

As comparison, we performed the STANDARD F COVID-19 Ag FIA test (SD BIOSENSOR Inc., Republic of Korea). It was a fluorescent immunoassay capable of detecting SARS-CoV-2 viral nucleoprotein antigens in human nasopharyngeal UTM swab specimens. Results were provided within 30 min and the observed values were expressed as a CutOff Index (COI) value, with COI  $\geq 1$  to be considered as sample positive for SARS-CoV-2 antigen.

IBM SPSS Statistics version 25 (manufactured by IBM in Armonk, NY, USA) was used to calculate 95 % confidence interval for sensitivity, specificity, negative and positive predictive values, and Cohen's kappa coefficient.

### 4. Results

Fiftyfive percent (60/110) of swabs tested positive in RT-qPCR while in the remaining 45 % (50/110) no virus was detected. Within the 60 positive samples, 23 had a Ct value for the N gene lower than 26, 32 a Ct ranging from 26 to 30 and the remaining 5 a Ct ranging from 31 and 35.

In Table 1, FRENDS™ COVID-19 Ag test results are shown in comparison with STANDARD F COVID-19 Ag FIA test and RT-qPCR results, for positive and negative samples, according Ct values for N gene and time from the onset of symptoms to sample obtention. With respect to RT-qPCR results, sensitivity, specificity, negative and positive predictive values of FRENDS™ COVID-19 Ag test were 93.3 % (95 % CI: 83.8–98.2), 100 % (95 % CI: 92.9–100), 92.5 % (95 % CI: 82.6–96.9) and 100 % (95 % CI: n.a.), respectively. Concordance between the two techniques was 96.3 % (Cohen's  $k = 0.93$ , 95 % CI: 0.86–0.99). As far as STANDARD F COVID-19 Ag FIA results, sensitivity, specificity, negative and positive predictive values were 86.7 % (95 % CI: 75.4–94.1), 100 % (95 % CI: 92.9–100), 86.0 % (95 % CI: 76.3–92.1) and 100 % (95 % CI: n.a.), respectively, compared with RT-PCR. FRENDS™ COVID-19 Ag test showed higher sensitivity both for samples with N gene Ct values from

**Table 1**

Sensitivity and specificity of FRENDS™ COVID-19 Ag test (NanoEntek, South Korea) and STANDARD F COVID-19 Ag FIA test (SD BIOSENSOR Inc., Republic of Korea), compared with RT-qPCR results. Swabs were divided into 3 groups based on RT-qPCR Ct values for N gene (<26, 26-30, 31-35) and into 2 groups according time from the onset of symptoms to sample obtention (<7 days, 7-14 days).

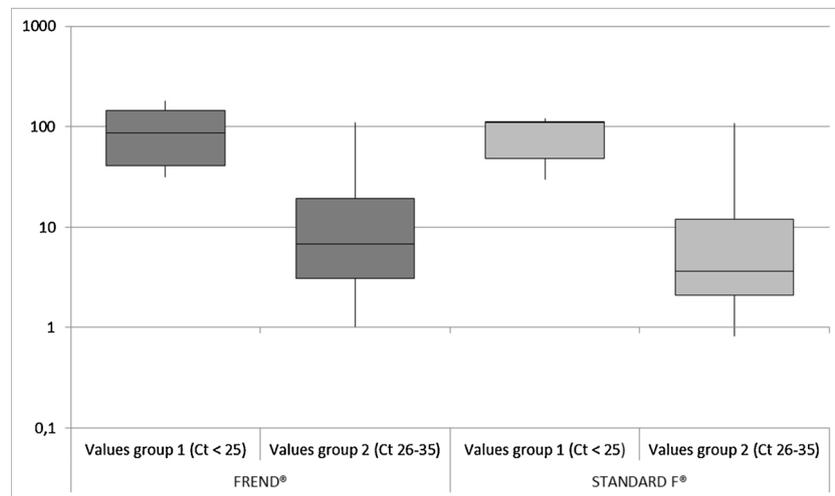
Swabs	FRENDS™ COVID-19 Ag		STANDARD F COVID-19 Ag FIA	
	Sensitivity(% 95 % C.I.)	Specificity (%, 95 % C. I.)	Sensitivity(% 95 % C.I.)	Specificity (%, 95 % C. I.)
RT-qPCR positive (n = 60)	93.3 (83.8–98.2)		86.7 (75.4–94.1)	
RT-qPCR negative (n = 50)		100 (92.9–100)		100 (92.9–100)
N gene Ct values				
Ct <26 (n = 23)	100 (85.2–100)		100 (85.2–100)	
Ct 26–30 (n = 32)	96.9 (83.8–99.9)		90.6 (75–98)	
Ct 31–35 (n = 5)	40 (5.3–85.3)		0 (0–52.1)	
Time from the onset of symptoms				
<7 days (n = 45)	95.6 (84.9–99.5)		88.9 (76–96.3)	
7–14 days (n = 15)	86.7 (59.5–98.3)		80 (51.9–95.7)	

26 to 35 and for samples collected up to 14 days after the onset of symptoms. Concordance between the two techniques was 92.7 % (Cohen's  $k = 0.86$ , 95 % CI: 0.76–0.95). The two Ag FIA assays showed a concordance of 96.4 % (Cohen's  $k = 0.93$ , 95 % CI: 0.86–0.99). In particular, for 4 samples (2 with a Ct value for N gene <30 and 2 with a Ct values from 31 to 35) there were discordant results between the two FIA methods. For these swabs, the mean value for FRENDS™ COVID-19 Ag test was 1.78, whereas for STANDARD F COVID-19 Ag FIA test was 0.88.

Fig. 1 compares numeric values provided by FRENDS™ COVID-19 Ag test and STANDARD F COVID-19 Ag FIA test for positive SARS-CoV-2 swabs in RT-qPCR. For positive samples with a RT-qPCR Ct value lower than 25, median value provided by FRENDS™ was 87.1 (interquartile range 40.8–146.0), whereas median COI provided by STANDARD F was 110.6 (interquartile range 48.2–112.6); as far as positive samples with a RT-qPCR Ct value ranging from 26 to 35, median value provided by FRENDS™ was 5.9 (interquartile range 2.1–18.3), whereas median COI provided by STANDARD F was 3.3 (interquartile range 1.7–11.6). With respect to time from the onset of symptoms to sample obtention, for positive samples collected up to 7 days after onset of symptoms median value provided by FRENDS™ was 30.2 (interquartile range 5.2–71.3), whereas median COI provided by STANDARD F was 15.2 (interquartile range 3.1–50.2); as far as positive samples collected from 7 to 14 days after onset of symptoms, median value provided by FRENDS™ was 7.6 (interquartile range 2.4–58.5), whereas median COI provided by STANDARD F was 6.2 (interquartile range 1.9–71.4).

### 5. Discussion

In the ongoing COVID-19 pandemic context, diagnostic testing for SARS-CoV-2 is crucial in order to limit the spread of the virus as well as appropriately manage infected patients. Different diagnostic test manufacturers have developed rapid tests based on SARS-CoV-2 proteins



**Fig. 1.** Box plot of numeric values provided by FRENDSM COVID-19 Ag test (NanoEntek, South Korea) and STANDARD F COVID-19 Ag FIA test (SD BIOSENSOR Inc., Republic of Korea) for positive SARS-CoV-2 swabs in RT-qPCR. A value  $\geq 1.00$  is considered positive for both assay. Group 1: positive samples with a RT-qPCR Ct value for N gene lower than 25. Group 2: positive samples with a RT-qPCR Ct value for N gene ranging from 26 to 35.

detection in respiratory samples. However, the analytical performances of these rapid antigenic tests depend on different factors including the viral load, the quality of the specimen and how it is processed.

In this study, we determined the performance characteristics of the FRENDSM COVID-19 Ag test for detecting SARS-CoV-2 virus in respiratory samples and compared the results with RT-qPCR as the gold standard and another FIA antigen test. Our data showed that FRENDSM COVID-19 Ag proved to be more sensitive and had several advantages such as the rapid answer in 3 min and the non-requirement of special equipment or personnel skills compared with molecular techniques.

The number of swabs collected and tested in this study could be a limit for our research; moreover, the Ct values of positive samples were never higher than 35. However, the study was performed in a “real world” clinical setting and within a few days, in order to evaluate the possible use of FRENDSM COVID-19 Ag as a screening tool in a large reference hospital.

Although negative results cannot rule out SARS-CoV-2 infection and made this test of little help when it is necessary to evaluate the progress of the disease and therefore, the state of recovery of the patient, our data suggest that FRENDSM system could be a useful device particularly in situations with limited access to molecular diagnostics and the need of rapid results in order to appropriately manage people and resources, i.e. Emergency Rooms or outpatient facilities.

In conclusion, FRENDSM COVID-19 Ag assay represents a valid frontline test for COVID-19 screening and it could ease the burden on the laboratories, reducing the time spent for diagnosis and the use of RT-qPCR.

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#### CRedit authorship contribution statement

**Andrea Orsi:** Writing - original draft, Formal analysis, Conceptualization. **Beatrice Marina Pennati:** Writing - original draft, Formal analysis, Conceptualization. **Bianca Bruzzone:** Conceptualization, Writing - review & editing, Supervision. **Valentina Ricucci:** Validation, Investigation, Data curation. **Diego Ferone:** Validation, Investigation. **Paolo Barbera:** Validation, Investigation. **Eleonora Arboscello:**

Validation, Investigation. **Chiara Dentone:** Validation, Investigation. **Giancarlo Icardi:** Conceptualization, Writing - review & editing, Supervision.

#### Declaration of Competing Interest

None.

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#### Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.jviromet.2021.114201>.

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